

# Effect of Chlordecone on Rate Brain Parts ACh Levels in Vivo

B. Hemavathi

*Lecturer in Zoology, Ramaraja College of Education, Tirupati, Chittoor District, Andhra Pradesh*

P.S.Poornima

*Lecturer in Zoology, S.V.Arts College Tirupati, Chittoor District, Andhra Pradesh.*

M.R.Rao

*Professor, Department of Zoology, S.V.University, Tirupati. Chittoor District, Andhra Pradesh.*

K.V.Kumari

*Professor, Department of Zoology, S.V.University, Tirupati. Chittoor District, Andhra Pradesh.*

**Abstract - Chlordecone belongs to Organochlorine group of pesticides and its use in agriculture is known to effect the normal functioning of non-target forms. We investigated the *IN VIVO* effect of Chlordecone on ACh levels of various brain parts of rat. Rats receiving a sub-lethal and lethal doses of Chlordecone showed increased the ACh content of all brain parts and the changes were found to be statistically significant over the control ( $P < 0.001$ ). The results indicate that Chlordecone treatment is favoring ACh accumulation in various regions of the brain.**

## I.INTRODUCTION

In the years which followed natural was good and concern was voiced from some quarters that the potential risks of pesticide out-weighed the benefits. This View has not diminished but had been tempered in recent years with the emergence of other concerns allied to food production and the realization that food like life is not risk free.

Recently, we've all been aware of the powerful forces at work that have impacted on our everyday lives and the livelihoods of those in agriculture. Those forces have been instrumental in setting the scene for the legislative frame work and consumer perception moveing forward. Many of us and in particular our farmers have been on the (BSE) roller coaster, experiencing constantly changing advice and confusion. The safety of genetically, modified crops have been under the spotlight with debate from all quarters ranging from pressure groups, through manufacturers and retailers to regulators across Europe .

### A. *Mode of action:*

The toxicity of organochlorides involves the functional disruption of sensory and motor nerve fibres and the motor cortex (Narahashi and Yamaski, 1968). The disruption of the neural mechanism is due to the penetration of organochlorides into the axon membrane altering the permeability to  $\text{Na}^+$  and  $\text{K}^+$ , subsequently inhibiting  $\text{Na}^+ - \text{K}^+$  and  $\text{Mg}^{2+}$  adenosine triphosphate activities in the nerve endings (Corbett, 1974).

Organochlorine compounds found to disrupt energy systems by inhibiting  $\text{Mg}^{2+}$  ATPase and aerobic segmental enzymes of brain (Cut Komp and Koch, 1981). Accumulation of ACh content in synaptic region was also noted (Omer et al., 1971). Inhibition of AChE and accumulation of ACh content by OCI compound shows the neurotoxic action (Omer et al., 1971).

### B. *Neurotoxic effects of chlordecone:*

Chlordecone, better known as Kepone®, belonging to OCI group of compounds, is a colourless, odorless crystalline solid, It is a polycyclic chlorinate compound with a caged structure similar to mirex (Desaiah, 1980). Several affected workers showed toxic symptoms involving primarily the nervous system, as evidenced by tremors, ataxia, slurred speech, mental changes, headache, irritability and exaggerated startle response (Desaiah, 1982; 1985). In addition to the neurotoxic symptoms, the chlordecone intoxicated workers also showed hepatosplenomegaly and presumed sterility (Guzelian, 1982).

The general biochemical toxic effects of chlordecone are summarized as below. The susceptibility of various animal species differ considerably although there is not much sex difference (Sherman and Ross, 1961; Gaines,

1969; Larson et al., 1979). In most studies, animals exposed to chlordecone show weight loss (Guzelian, 1982). Chlordecone was a neurotoxin, estrogen, hepatotoxin and potentiator of halomethane toxicity.

## II. MATERIALS AND METHODS

Albino rats of the weight range  $150 \pm 5$  g were used for the present study. Rats were maintained at constant temperature of  $25 \pm 5$  °C, humidity 60-70% they were fed with ad libitum commercial diet supplied by Sri Kamadhenu Agencies, Bangalore, India. 24 hrs prior to experiment they were tested with free access to water. They were fasted with free access to water. They were divided into groups of 10 each, maintained in separate cages and were used either for LD50 study or chlordecone treatment. For sub-lethal and lethal doses of chlordecone treatment the numbers of rats used were seven each for every treatment.

After determination of LD<sub>50</sub> dose of chlordecone to rat per 48 hours, a rat colony was divided into three groups of seven each. I group acted as control one, the II group was gavaged with a sub-lethal chlordecone dose of 29.71 mg/kg and the III with a lethal dose chlordecone of 89.13 mg/kg. after 48 hrs chlordecone treatment of rats, the control and experimental group of rats individually were anaesthetized with Katamine (7mg/Kg).

From the whole brains dissected, the various brain parts like cerebellum, hypothalamus, midbrain, striatum, hippocampus, cerebral cortex and medulla oblongata were separated and individually frozen in liquid nitrogen and were stored at -80°C till used.

## III. METHODS

### *Determination of Acetylcholine Content (ACh)*

ACh content in control and experimental samples was estimated by the method of Hestrin as described by Augustinsson (1957). The tissues (different parts of rat brain regions) individually were isolated, sliced, weighed and were transferred into clean test tubes. The tubes were kept in boiling water bath for 5 minutes to inactivate the acetylcholinesterase (AChE) enzyme activity and to release bound ACh as described by Vasantha et al., (1975) The tubes were cooled and the contents were homogenized in 2.0ml of distilled water, 2.0 ml of alkaline hydroxylamine hydrochloride and 1.0 ml of hydrochloric acid (1:1-HCl : H<sub>2</sub>O) were added to the homogenates. The contents were centrifuged at 2000 g/10 min and 1.0 ml of ferric chloride (FeCl<sub>3</sub>) was added to the supernatant. The optical density of the sample was measured at 540 nm in a spectrophotometer using the reagent blank ACh content was expressed as  $\mu$ moles of ACh /gm wet wt of tissue.

## IV. RESULTS AND DISCUSSION

### *RESULTS:*

The data presented in table-1 shows the ACh content of various parts of control and chlordecone treated rat brain. A sub-lethal and lethal dose of chlordecone has significantly ( $P < 0.001$ ) enhanced the rat cerebellum, Hypothalamus, mid brain, striatum, hippocampus, cerebral cortex and medulla oblongata ACh content. Concerning to the percent changes LD<sub>50</sub> dose chlordecone treated rat brain parts showed more percent increase of their ACh content over the control compared to  $\frac{1}{3}$  LD<sub>50</sub> dose chlordecone treated ones (fig-1). The results of table-1 further depicts that in the control group of brain parts the tissue specific trend for ACh observed was more for striatum and was followed by medulla oblongata > hippocampus > hypothalamus > mid brain > cerebral cortex > Cerebellum.

### *Discussion:*

Acetylation of the nitrogenous alcohol choline gives rise to ACh, now recognized as a neurotransmitter in all major groups of animals (Hoar, 1976). The biosynthesis is catalyzed by choline acetylase and readily reversed by acetylcholinesterase (AChE).

The most widely recognised action sites of ACh are the endings of vertebrate motor nerves, the endings of the autonomic preganglionics and the parasympathetic post ganglionics. ACh is concerned with various visceral functions in both invertebrates and vertebrates.

As given by Hoar (1976), Claude Bernard launched the history of transmitter pharmacology in 1857 when he discovered that nerve stimulation failed to excite muscles that were poisoned with a plant alkaloid curare, even through the muscle remained responsive to direct stimulation.

The physiologists at present have discovered a wide range of drugs that interact with ACh. Hemicholinium - 3 prevents ACh synthesis in motor nerves, botulinum toxin, one of the most potent poisons in the world, prevent its release. Several different substances termed AGONISTS, act like ACh on the post-synaptic membrane; well known examples are carbachol, succinylcholine, necamethenium and nictene.

Biochemical experiments have indicated that exogenously applied NO is able to influence the release of several neurotransmitters such as glutamate, GABA, dopamine, ACh, adrenalin (Garth waite and Boulton

1995. The definitive role of ACh in regulation of NOS functions is well documented by Sudhir et al., (1994). He revealed that ACh -induced vasodilation is mediated by the release of endothelium-derived relaxing factors (EDRF's) (Furchgott and Zawadzki, 1980) one of which is NO (Palmer et al., 1987; Ignarro et al., 1987). Endothelium-dependent vasorelaxation influences coronary blood flow and vascular resistance through effects on both epicardial and resistance coronary arteries (Luscher et al., 1990). The role of ACh in NO release at least in the cardiovascular system is reported by many investigators (Furchgott, 1993; Vanhoutt; 1993, Busse et al., 1993; Sudhir et al., 1994). In view of the key role played by the neurotransmitter molecule like ACh in NO release, the author attempted to study the in vivo effect of chlordecone on ACh levels of various brain parts of rat.

A sub-lethal and lethal dose of chlordecone over a period of 48 hours increased the ACh content of all brain parts studied in the present investigation (table-1) and the changes were found to be statistically significant over the control ( $P < 0.001$ ). The results indicate that chlordecone treatment is favouring ACh accumulation in various regions of the brain.

The toxicity of OCI compounds involve the functional disruption of sensory and motor nerve fibres (Narahashi and Yamaski, 1968). Accumulation of ACh content in synaptic region was noted by Omer et al., (1971). Inhibition of AChE and accumulation of ACh content by OCI compounds shows their neurotoxic effect (Omer et al., 1971). ACh has been identified in many areas of the brain, and in the opinion of some, it is the only component that fits the most stringent criteria for a CNS transmitter (Myers, 1974). ACh is released from the endings of cholinergic fibres to effect synaptic transmission. Under normal circumstances ACh is hydrolysed by AChE immediately. There is no accumulation of ester as well. So the normal function of ACh depends upon its rapid destruction by AChE Maslora(1981).

Cholinesterases are of two types, viz. a) true cholinesterases and b) pseudocholinesterases or nonspecific cholinesterases. They differ in substrate preference and specificity, kinetics, substrate hydrolysis and sensitivity to some inhibitors in different animal systems (Augustinsson, 1963; Perse, 1972; Silver, 1974).

The physiological role of pseudocholinesterases has not yet been clearly specified. The pseudocholinesterases are relatively less efficient than AChE at low concentrations of substrate but more efficient at higher concentrations. Butyrylcholinesterase (BuChE) is a less specialized enzyme than AChE. Unlike AChE, it lacks anionic site in a position that specifically adapts it to react with ACh. It does, however, slowly react with ACh. It also reacts with a wide range of esters (Ecobichon and Comean, 1973).

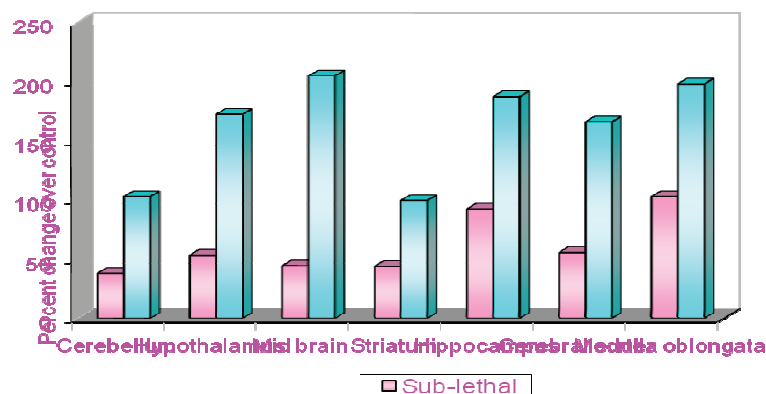
Cholinesterases and ACh are found to differ in their concentrations in the different areas of rat brain. Striatum is the area rich in cholinergic neurons and the levels of AChE and ACh are high in this area (Stavinoha et al., 1976; Ehlert et al., 1980a; 1980b; ) and lowest activity of AChE and ACh were in the area of cerebellum (Bennet et al., 1966). Studies on different brain areas where functional deficits should be readily associated and recognized are of importance for a fuller understanding of the toxic actions of insecticides (Shih, 1982). The functions of brain regions also significantly vary from each other. In view of this, a study of the response of different brain regions to chlordecone toxicity with reference to ACh content would be worth while.

From the data presented in table-1, it can be inferred that the control rat striatum showed highest content of its ACh and was followed by medulla oblongata > hippocampus > hypothalamus > midbrain > cerebral cortex > cerebellum and the trends obtained were in agreement with the reports of Bennet et al., (1966)

The percent elevation of ACh for any given brain region was more in lethal dose chlordecone treated brain regions, inferring that higher the dose of chlordecone higher the percent elevation of ACh (table-1).

The sum total of the cholinergic substances obtained by the assay presented for ACh determination be construed as the amount of ACh present or have accumulated in the respective tissues during chlordecone exposure could be due to lesser breakdown of ACh because of AChE is known to be inhibited upon OCI compounds exposure in animal models (Omer et al., 1971).

A reduction in muscarinic receptor density by chronic administration of OI like OP's is closely related to the degree of ChE inhibition (Sivam et al., 1983., Yamada et al., 1983a). The state of subsensitivity is the result of AChE inhibition and increased accumulation of ACh to which effector cells are exposed, causing the responsiveness of the cells to decrease (Foley and Mc Phillips 1973). It was reported that OI exposure result in decrease in the density of muscarinic receptors and this occurs when the brain AChE is inhibited (Russell et al., 1975). OI's further were shown to cause down-regulation or subsensitivity of cholinergic receptors resulting from elevated ACh concentration (Costa et al., 1982) as was observed in the present investigation. Number of environmental toxicants have been shown to impair ACh functioning including receptor mediated events in neuronal systems (Buckingham et al., 1997; Chao et al., 1997; Xu Hai-Bin et al., 1997; Katz et al., 1997; Guilhermino et al., 1998; Matsuda et al., 1998; Stefanidou et al., 1998; Tang et al., 1998; Ostby et al., 1999). Based on the above, the elevation in brain regions ACh levels in sub-lethal and lethal dose chlordecone treated rats (table-1) and blockade of receptor functions by OI's as cited above may inhibit/reduce the binding of ACh to its receptor sites (Van Helden et al., 1998; Slotkin 2009; Shelton, 2012) and thus in the current study one of the OI's studied chlordecone may impair the usual functioning of rat brain based ACh.

**Fig.1: Percent change of sub-lethal and lethal**Table-1 :Effect of sub-lethal and lethal doses of Chlordecone on ACh levels in various parts of rat brain *in vivo*.(Values expressed as  $\mu\text{M}$  of ACh/gm wet wt of tissue)

Name of the Brain part	Control	Sub-lethal Experimental	Lethal Experimental
Cerebellum SD PC t	0.482 "0.020	0.664 "0.033 37.75 P<0.001	0.978 "0.024 102.90 P<0.001
Hypothalamus SD PC t	0.602 "0.041	0.923 "0.027 53.32 P<0.001	1.64 "0.044 172.42 P<0.001
Mid brain SD PC t	0.584 "0.021	0.844 "0.032 44.52 P<0.001	1.78 "0.062 204.79 P<0.001
Striatum SD PC t	1.34 "0.071	1.93 "0.044 44.02 P<0.001	2.67 "0.057 99.25 P<0.001
Hippocampus SD PC t	0.821 "0.037	1.58 "0.062 92.44 P<0.001	2.36 "0.091 187.45 P<0.001
Cerebral cortex SD PC t	0.541 "0.046	0.841 "0.037 55.45 P<0.001	1.44 "0.042 166.17 P<0.001
Medulla oblongata SD PC t	0.917 "0.037	1.86 "0.036 102.83 P<0.001	2.73 "0.075 197.70 P<0.001

Each value is the mean  $\pm$  SD of 7 samples.

SD : Standard deviation

PC : Percent change over control ones

## REFERENCES

- [1] Aldous, C.N., Chetty, C.S., Mehendale, H.M. and Desai, D. 1984. Lack of effects of chlordecone on synthesis rates steady state levels and metabolites of catecholamines in rat brain. *Neurotoxicity*, **5(2)**: 59.
- [2] Augustinsson, K.B. 1957. In *Methods of biochemical analysis*, vol.5 (ed. Glick, D). *Inter Science Publishers Inc.*, New York, USA, pp.1
- [3] Augustinsson, K.B. 1963. Cholinesterases and anticholinesterase agents, Ed. G.B. Koelle. Springer Verlag, Berlin, 89-128.
- [4] Bennet E.L., Diamond, M.C., Morimoto, H. and Herbert, M. 1966. AChE activity and weight measures in fifteen brain areas from six lines of rats. *J. Neurochem.*, **13**: 563-572.
- [5] Buckingham, S.D., Lapied, B., Lecorronc, H., Grolleau, F. and Sattelle, D.B. 1997. Imidachloprid actions on insect neuronal acetylcholine receptors. *J. Exp. Biol.*, Nov; **200 (21)**: 2685-2692.
- [6] Busse, R., Mulsch, A., Fleming, I. and Hecker, M. 1993. Mechanisms of Nitric Oxide Release from the Vascular Endothelium. *Circulation*, **87 (Suppl v)**: 18-25.
- [7] Chao, Shirley Lee and John E., Casida. 1997. Interaction of imidachloprid metabolites and analogs with the nicotinic acetylcholine receptor of mouse brain in relation to toxicity. *Pesticide 1, Biochem. Physiol.*, May; **58(1)**: 77-88.
- [8] Corbett, J.R. 1974. In the biochemical mode of action of Pesticides. Academic Press, pp. 330.
- [9] Costa, L.G., Schwab, B.W. and Murphy, S.D. 1982. Tolerance to anticholinesterase compounds in mammals. *Toxicology*, **25**: 79-97.
- [10] Cutkomp, L.K. and Koch, R.B. 1981. Inhibition of ATPase by chlorinated hydrocarbons. Presented in the symposium on mode of action of chlorinated hydrocarbons. Personal Communication.
- [11] Desai, D. 1980. Comparative effects of chlordecone and mirex on rat cardiac ATPases and binding of  $^3\text{H}$ -catecholamines. *J. Environ. Pathol. Toxicol.*, **4**: 237-248.
- [12] Desai, D. 1982. Effect of chlordecone on membrane receptors in brain. In: *Proceedings of the International Conference on Neurotoxicology of Selected Chemicals. Chlordecone Symposium* J.M. Cranmer, D. Desai, H. A. Tilson, eds. Little Rock, Intox Press.
- [13] Desai, D. 1985. Chlordecone interaction with catecholamine binding and uptake in rat brain synaptosomes. *Neurotoxicol.*, **6(1)**: 159-166.
- [14] Ecobichon, D.J. and Comean, A.M. 1973. Pseudocholinesterases of mammalian plasma. *Toxicol. Appl. Pharmacol.*, **24**: 92-100.
- [15] Ehler, F.J., Kolska, N. and Fairhurst, A.S. 1980a. Altered [ $^3\text{H}$ ]-quinuclidinyl benzilate binding in the striatum of rats following cholinesterase inhibition with diisopropyl fluorophosphate. *Mol. Pharmacol.*, **17**: 24-30.
- [16] Ehler, F.J., Dumont, Y., Roeske, W.R. and Yamamura, H.I. 1980b. Muscarinic receptor binding in rat brain using the agonist [ $^3\text{H}$ ] cis-methylidioxolane. *Life Sci.*, **26**: 961-967.
- [17] Foley, D.J. and McPhillips, J.J. 1973. Response of the rat ileum, uterus and vas deferens to carbachol and acetylcholine following repeated daily administration of cholinesterase inhibitor. *Br. J. Pharmacol.*, **48**: 418-425.
- [18] Furchgott, R.F. and Zawadzki, J.V. 1980. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature*, **288**: 373-376.
- [19] Furchgott, R.F. 1993. The discovery of Endothelium-Dependent Relaxation. *Circulation*, **87 (Suppl V)**: 3-8.
- [20] Gaines, T.B. 1969. Acute toxicity of pesticides. *Toxicol. Appl. Pharmacol.*, **14**: 515-534.
- [21] Garthwaite, J. and Boulton, L.L. 1995. Nitric oxide signaling in the central nervous system. *Annu. Rev. Physiol.*, **57**: 683.
- [22] Guzelian, P.S. 1982. Comparative toxicology of chlordecone (Kepone) in humans and experimental animals. *Ann. Rev. Pharmacol. Toxicol.*, **22**: 89-113.
- [23] Guilhermino, Lucia, Amadeu M.V.M., Soares, Arselio, P., Carvalho, and Celeste Lopes, M. 1998. Correlation between whole blood cholinesterase activity and cerebral cortex cholinesterase activity in rats treated with parathion. *Chemosphere*, Sept; **37 (7)**: 1385-1393.
- [24] Hoar, W.S. 1976. "General and comparative Physiology" 2<sup>nd</sup> edition, Prentice Hall of India. Hedged Lourent. 1973. In *Environmental Pollution*. Hart Rinchart and; Winston, New York.
- [25] Ignarro, L.J., Byrns, R.E., Buga, G.M. and Wood, K.S. 1987. Endothelium-derived relaxing factor from pulmonary artery and vein possess pharmacological and chemical properties identical to those of nitric oxide radical. *Cir. Res.*, **61**: 866-879.
- [26] Katz, Elizabeth, J., Vania, I., Cortes, Mohyee, E., Eldegrawi, And Amira, T., Eldefrawi. 1997. Chlorpyrifor, parathion and their oxons bind to and desensitize a nicotinic acetylcholine receptor. *Toxicol. Appl. Pharmacol.*, Oct; **146 (2)**: 227-236.
- [27] Larson, P.S., Egle, J.L., Henninger, G.R., Lane, F.W. and Borzelleca, J.F. 1979. Acute, subchronic and chronic toxicity of chlordecone. *Toxicol. Appl. Pharmacol.*, **48**: 29-41.
- [28] Luscher, T.F., Richard, V., Tschudi, M., Yang, Z. and Boulanger, C. 1990. Endothelial control of vascular tone in large and small coronary arteries. *J. Am. Coll. Cardiol.*, **15**: 519-27.
- [29] Maslora, O.Y. 1981. Cholinesterase activity in estuarine fish accumulating DDT. *Gidrobiol. Zh.*, **17**: 79.
- [30] Matsuda, K., Buckingham, S.D., Freman, J.C., Squire, M.D., Baylis, H.A. and Sattelle, D.B. 1998. Effects of the  $\alpha$  subunit on imidachloprid sensitivity of recombinant nicotinic acetylcholine receptors. *Br. J. Pharmacol.*, Feb; **123(3)**: 518-524.
- [31] Myers, R.D. 1974. *Handbook of Drug and chemical stimulation of the Brain*: In: Behavioural pharmacological and physiological Aspects. Van Nostrand Reinhold, New York.
- [32] Narahashi and Yamaski. 1986. Mechanism of increase in negative after potential by dicophanum in the giant axon of the Cockroach. *J. Physiol.*, (London). **152**: 122.
- [33] Omer, S., Vincent, V. and Ecobichon, D.J. 1971. Biochemical effects of chlorinated insecticides DDT and Dieldrin by Kohll et al., 1975 in: *Isr Ind. Res.* **34**: 469.
- [34] Ostby, Joseph, William, R., Kelce, Christy Lambright, Cynthia, J., Wolf, Peter Mann and Earl Gray, L. Jr. 1999. The fungicide procymidone alters sexual differentiation in the male rat by acting as an androgen receptor antagonist *In vivo* and *in vitro*. *Toxicol. Indus. Health*, Jan - March; **15(1-2)**: 80-93.

- [35] Palmer, R.M.J., Ferrige, A.G. and Moncada, S. 1987. Nitric oxide release accounts for the biological activity of endothelium – derived relaxing factor. *Nature*, **327** : 524-526.
- [36] Perse, A.G.E. 1972. Histochemistry. Theoretical and applied. Churchill. Living stone, Edn Edinburgh. 2.
- [37] Russell, R.W., Overstreet, D.H., Cotman, C.W., Carson, V.G., Churchill, L., Dalglish, F.W. and Vasquez, B.J. 1975. Experimental tests of hypothesis about neurochemical mechanisms underlying behavioural tolerance to the anticholinesterase Diisopropyl fluorophosphate. *J. Pharmacol. Exp. Ther.*, **192** : 73-85.
- [38] Sherman, M. and Ross, E. 1961. Acute and subacute toxicity of insecticides to chicks. *Toxicol. Appl. Pharmacol.*, **3** : 521-533.
- [39] Shih, T.M. 1982. Time course of soman on acetylcholine and choline levels in six discrete areas of the rat brain. *Psychopharmacologia*, **78** : 170-175.
- [40] Shelton, J.F. 2012 Tipping the Balance of Autism Risk: potential Mechanisms linking. *Environ. Health. perspect*, V.120(7): Jul.
- a. Silver, A. 1974. The biology of cholinesterases. North Holl and, Amsterdam. 36.
- [41] Sivam, S.P., Norris, J. C., Lim, D.K., Hoskins, B. and Ho, I.K. 1983. Effect of acute and chronic cholinesterase inhibition with Diisopropyl fluorophosphate on muscarinic, dopamine and GABA receptors of the rat striatum. *J. Neurochem.*, **40** : 1414-1422.
- [42] Slotkin, T.A. 2009. Consumption of a High-fat diet in adulthood Ameliorates the effects. *Environ. Health. perspect*, V.117 (6): Jun.
- [43] Stavinoha, W.B., Modak, A.T. and Weintraub, S. T. 1976. Rate of accumulation of acetylcholine in discrete regions of the rat brain after Dichlorvos treatment. *J. Neurochem.*, **27** : 1375 – 1378.
- [44] Stefanidou, M., Pappas, F., Methenitou, G., Dona, A., Alevisopoulos, G. and Koutselis, A. 1998. Bee pseudo-cholinesterase as an indicator of exposure to anticholinesterase insecticides. *Veterinary and Human Toxicology*, **40 (6)** ; 326-327.
- [45] Sudhir, K., MacGregor, J.S., Amidon, T.M., Gupta, M., Yock, P.G. and Chatterjee, K. 1994. Differential contribution of nitric oxide to regulation of vascular tone in coronary conductance and resistance arteries: Intravascular ultrasound studies. *Am. Heart. J.*, **127** : 858-865.
- [46] Tang, Hai-Wang and Gudrun Cassel. 1998. Effect of soman on N-methyl-D-aspartate-stimulated [<sup>3</sup>H] norepinephrine release from rat cortical slices. *Toxicol. Lett.*, (Shannon) Nov – 12; **99(3)** : 169-173.
- [47] Van Helden, Herman, P.M., Bas Groen, Eyten Moor, Ben H.C., Westerink and Piet, L.B., Bruijnzeel. 1998. New generic approach to the treatment of organophosphate poisoning : Adenosine receptor mediated inhibition of ach-release. *Drug and Chemical Toxicology an International Journal for Rapid Communication*, **21 (Suppl.1)** : 171-181.
- [48] Vanhoutte, P.M. 1993. Other endothelium-derived vasoactive factors. *Circulation*, **87 (Suppl V)** : 9-17.
- [49] Vasanta, N., Venkatachari, S.A.T., Muralinohan, P. and Sasira Babu, K. 1975. "On the acetylcholine contents in the *Scorpion, Heterometrus fulvipes* C.Koch: *Experientia*, **31** : 451-452.
- [50] Xu Hai – Bin, He xi – Wen, Xiezuoping and He Fengsheng. 1997. Blocking effects of Omethoate on acetylcholine receptor channels. *Zhongguo Yaolixue Yudilixue Zazhi*. Nov; 11 (4) :286 – 290.
- [51] Yamada, S., Isogai, M., Okudaira, H. and Hayashi, E. 1983a. Regional adaptation of muscarine receptors and choline uptake in brain following repeated administration of Diisopropyl fluorophosphate and atropine. *Brain Res.*, **268** : 315-320.